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## INTERNATIONAL APPLICATION PUBLISHED UNDER THE PATENT COOPERATION TREATY (PCT)

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<b>(21) International Application Number:</b> PCT/US92/05788 <b>(22) International Filing Date:</b> 10 July 1992 (10.07.92)  <b>(30) Priority data:</b> 728,093 10 July 1991 (10.07.91) US 728,194 10 July 1991 (10.07.91) US  <b>(71) Applicant:</b> IGEN, INC. [US/US]; 1530 East Jefferson Street, Rockville, MD 20852 (US).  <b>(72) Inventors:</b> MASSEY, Richard, J. ; 5 Valerian Court, Rockville, MD 20852 (US). BLACKBURN, Gary, F. ; 23627 Rolling Fork Way, Gaithersburg, MD 20882 (US). WILKINS, Elizabeth, W. ; 20414 Ambassador Terrace, Germantown, MD 20874 (US). SHAH, Haresh, P. ; 897 Clopper Road, T4, Gaithersburg, MD 20878 (US). LE-LAND, Jonathan, K. ; 14002 Chestnut Court, Laurel, MD 20707 (US).		<b>(74) Agents:</b> EVANS, Barry et al.; Curtis, Morris & Safford, 530 Fifth Avenue, New York, NY 10036 (US).  <b>(81) Designated States:</b> AU, CA, JP, KR, European patent (AT, BE, CH, DE, DK, ES, FR, GB, GR, IT, LU, MC, NL, SE).  <b>Published</b> <i>With international search report.</i>
<b>(54) Title:</b> METHODS AND APPARATUS FOR IMPROVED LUMINESCENCE ASSAYS USING PARTICLE CONCENTRATION AND CHEMILUMINESCENCE DETECTION  <b>(57) Abstract</b>  What is described are methods and apparatus for performing a binding assay for an analyte of interest present in a sample. The methods include the steps of: forming a composition containing the sample, an assay-performance-substance which contains a component linked to a label compound capable of chemiluminescing when triggered, and a plurality of particles capable of specifically binding with the analyte and/or the assay-performance-substance; incubating the composition to form a complex which includes a particle labeled component; collecting the complex in a collection zone; introducing into the collection zone a trigger capable of triggering the label such that the label luminesces; and measuring the emitted luminescence to measure the presence of the analyte of interest in the sample.		

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**Methods and Apparatus for Improved Luminescence Assays  
Using Particle Concentration and Chemiluminescence Detection**

This application is a continuation-in-part of  
copen ding Massey et al. application Ser. No. 07/652,427  
5 filed February 6, 1991, which is a continuation-in-part of  
application Ser. No. 07/539,389 filed June 18, 1990, which  
is a continuation of application Ser. No. 07/266,882 filed  
November 3, 1988, now abandoned, entitled  
Electrochemiluminescent Assays, and this application is a  
10 continuation-in-part of copending application Ser. No.  
07/539,389 filed June 18, 1990, which is a continuation of  
application Ser. No. 07/266,882 filed November 3, 1988, now  
abandoned, entitled Electrochemiluminescent Assays. The  
subject matter of this application is incorporated herein by  
15 reference.

**Field of The Invention**

This application relates generally to methods and  
apparatus for conducting binding assays, more particularly  
to those which measure the presence of an analyte of  
20 interest by measuring luminescence emitted by one or more  
labeled components of the assay system. More specifically,  
the invention relates to precise, reproducible, accurate  
homogeneous or heterogeneous specific binding assays of  
improved sensitivity in which the luminescent component is  
25 concentrated in the assay composition and collected before  
being caused to electrochemiluminescence.

**Background of The Invention**

Numerous methods and systems have been developed  
for the detection and quantitation of analytes of interest  
30 in biochemical and biological substances. Methods and  
systems which are capable of measuring trace amounts of  
microorganisms, pharmaceuticals, hormones, viruses, anti-  
bodies, nucleic acids and other proteins are of great value  
to researchers and clinicians.

35 A very substantial body of art has been developed  
based upon the well known binding reactions, e.g., antigen-  
antibody reactions, nucleic acid hybridization techniques,  
and protein-ligand systems. The high degree of specificity

in many biochemical and biological binding systems has led to many assay methods and systems of value in research and diagnostics. Typically, the existence of an analyte of interest is indicated by the presence or absence of an observable "label" attached to one or more of the binding materials. Of particular interest are labels which can be made to luminesce through photochemical, chemical, and electrochemical means. "Photoluminescence" is the process whereby a material is induced to luminesce when it absorbs electromagnetic radiation. Fluorescence and phosphorescence are types of photoluminescence. "Chemiluminescent" processes entail the creation of luminescent species by chemical transfer of energy. "Electrochemiluminescence" entails creation of luminescent species electrochemically.

Chemiluminescent assay techniques where a sample containing an analyte of interest is mixed with a reactant labeled with a chemiluminescent label have been developed. The reactive mixture is incubated and some portion of the labeled reactant binds to the analyte. After incubation, the bound and unbound fractions of the mixture are separated and the concentration of the label in either or both fractions can be determined by chemiluminescent techniques. The level of chemiluminescence determined in one or both fractions indicates the amount of analyte of interest in the biological sample.

The incubated sample may be exposed to a voltammetric working electrode in order to trigger luminescence. In the proper chemical environment, such electrochemiluminescence is triggered by a voltage impressed on the working electrode at a particular time and in a particular manner. The light produced by the label is measured and indicates the presence or quantity of the analyte. For a fuller description of such electrochemiluminescent techniques, reference is made to PCT published application US85/01253 (WO86/02734), PCT published application number US87/00987, and PCT published application US88/03947. The disclosures of the aforesaid applications are incorporated by reference.

It is desirable to carry out chemiluminescent assays without the need for a separation step during the assay procedure and to maximize the signal modulation at different concentrations of analyte so that precise and sensitive measurements can be made.

Among prior art methods for separation assays are those such as described in U.S. Patent No. 4,141,687 and European Patent Application 0,030,087 which relate to magnetically separating particles in a conduit after which the particles are removed to a separate chamber for analysis of the label.

Among prior art methods for nonseparation assays are those which employ microparticulate matter suspended in the assay sample to bind one or more of the binding components of the assay.

U.S. Patent No. 4,305,925 relates to the detection and determination of clinically relevant proteins and peptides by means of nephelometric and turbidimetric methods. The methods disclosed involve binding the antigen or antibody to latex particles which perform the function of light scattering or adsorption.

U.S. Patent No. 4,480,042 relates to techniques employing particle reagents consisting of shell-core particles. The shell contains functional groups to which compounds of biological interest can be covalently bonded, and the high refractive index of the core results in high sensitivity to light scattering measurements. The technique is based upon agglutination reactions which result from the reaction of bivalent antibodies with multivalent antigens of interest to produce aggregates which can be detected and/or measured in various ways.

U.S. Patent No. 4,419,453 likewise relates to the use of colored latex agglutination test methods useful for detecting the presence of immunochemicals such as antibodies and immunogens.

Based upon this prior art, it would not have appeared possible to use microparticulate matter in assays wherein a luminescent phenomenon is measured. One would

expect that the luminescence from free chemiluminescent moieties would be absorbed, scattered, or otherwise suffer interference from the microparticulate matter.

Contrary to that expectation, U.S. Application  
5 Serial 266,882 (PCT published application U.S. 89/04919)  
teaches sensitive, specific binding assay methods based on a  
luminescent phenomenon wherein inert microparticulate matter  
is specifically bound to one of the binding reactants of the  
assay system. The assays may be performed in a  
10 heterogeneous (one or more separation steps) assay format  
and may be used most advantageously in a homogeneous  
(nonseparation) assay format.

US89/04919 relates to a composition for an assay  
based upon a binding reaction for the measurement of  
15 luminescent phenomenon, which composition includes a  
plurality of suspended particles having a surface capable of  
binding to a component of the assay mixture. In another  
aspect, it is directed to a system for detecting or  
quantitating an analyte of interest in a sample, which  
20 system is capable of conducting the assay methods using the  
assay compositions of the inventions. The system includes  
means for inducing the label compound in the assay medium to  
luminesce, and means for measuring the luminescence to  
detect the presence of the analyte of interest in the  
25 sample.

Thus, US89/04919 is directed to methods for the  
detection of an analyte of interest in a sample, which  
method includes the steps of (1) forming a composition  
comprising (a) a sample suspected of containing an analyte  
30 of interest, (b) an assay-performance-substance selected  
from the group consisting of (i) analyte of interest or  
analog of the analyte of interest, (ii) a binding partner of  
the analyte of interest or its said analog, and (iii) a re-  
active component capable of binding with (i) or (ii),  
35 wherein one of said substances is linked to a label compound  
having a chemical moiety capable of being induced to  
luminesce, and (c) a plurality of suspended particles  
capable of specifically binding with the analyte and/or a

substance defined in (b)(i), (ii), or (iii); (2) incubating the composition to form a complex which includes a particle and said label compound; (3) inducing the label compound to luminesce; and (4) measuring the luminescence emitted by the composition to detect the presence of the analyte of interest in the sample. Those same methods may be used to quantify the amount of analyte in a sample by comparing the luminescence of the assay composition to the luminescence of a composition containing a known amount of analyte.

10           Analogues of the analyte of interest, which may be natural or synthetic, are compounds which have binding properties comparable to the analyte, but include compounds of higher or lower binding capability as well. Binding partners suitable for use in the present invention are well-  
15 known. Examples are antibodies, enzymes, nucleic acids, lectins, cofactors and receptors. The reactive components capable of binding with the analyte or its analog and/or with a binding partner thereof may be a second antibody or a protein such as Protein A or Protein G or may be avidin or  
20 biotin or another component known in the art to enter into binding reactions.

Advantageously, the luminescence arises from electrochemiluminescence (ECL) induced by exposing the label compound, whether bound or unbound to specific binding  
25 partners, to a voltammetric working electrode. The ECL reactive mixture is controllably triggered to emit light by a voltage impressed on the working electrode at a particular time and in a particular manner to generate light. Although the emission of visible light is an advantageous feature the  
30 composition or system may emit other types of electromagnetic radiation, such as infrared or ultraviolet light, X-rays, microwaves, etc. Use of the terms "electrochemiluminescence," "electrochemiluminescent" "luminescence," "luminescent," and "luminesce" includes the emission of  
35 light and other forms of electromagnetic radiation.

The methods taught in US89/04919 permit the detection and quantitation of extremely small quantities of analytes in a variety of assays performed in research and

clinical settings. The demands of researchers and clinicians makes it imperative, however, to lower the detection limits of assays performed by these methods to increase the sensitivities of those assays and to increase the speed at which they can be performed.

Various methods are known in the art for increasing the signal from labeled species by concentrating them before subjecting them to a measurement step. In U.S. Patent No. 4,652,333, for example, particles labeled with fluorescent, phosphorescent or atomic fluorescent labels are concentrated by microfiltration before a measurement step is performed.

It is also known in the art to concentrate labeled immunochemical species prior to a measurement step, by, e.g., drawing magnetically responsive labeled particles to the surface of a measurement vessel. In U.S. Patents No. 4,731,337, 4,777,145, and 4,115,535, for example, such particles are drawn to the vessel wall and then are irradiated to excite a fluorophoric emission of light.

In U.S. Patent No. 4,945,045, particles are concentrated on a magnetic electrode. An electrochemical reaction takes place at the electrode facilitated by a labeled chemical mediator. The immunochemical binding reaction alters the efficiency of the mediator resulting in a modulated signal when binding takes place.

#### Objects of The Invention

It is therefore a primary object of this invention to provide homogeneous (non-separation) and heterogeneous (separation) methods, reagents and apparatus, for the conduct of binding assays.

It is a further object of this invention to provide non-separation, specific bonding assays, reagents and apparatus, based upon the measurement of electrochemiluminescence emitted from an assay composition containing microparticulate matter.

It is a further and related object to provide such assays, reagents and apparatus having improved sensitivity,

faster assay time, greater sensitivity, lower detection limits and greater precision than has heretofore been achieved.

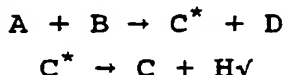
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## Description of the Invention

### Definition of Terms

Chemiluminescence is defined as a luminescence reaction in which the energy responsible for generating the high-energy excited state of a molecule is derived from an  
10 energetic chemical reaction. A chemiluminescent reaction thus involves the direct conversion of chemical energy to electromagnetic radiation (ultraviolet, visible, or infrared radiation). Luminescence occurs when the excited-state molecule returns to its ground-state energy level, emitting  
15 a photon having a particular wavelength which is characteristic of the molecule and the energy of its excited state relative to its ground-state.

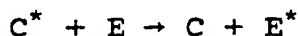
Energy is generated by many chemical reactions; such reactions are called exothermic reactions. In most  
20 cases the energy appears as heat and induces vibrational, rotational, and translational energy in the molecule. In a chemiluminescence reaction at least part of this energy is channeled into the formation in the high-energy excited state. This generally requires a highly energetic and rapid  
25 reaction of two molecules, one of which is capable of luminescence emission:



The quantity  $h\nu$  represents a photon of  
30 electromagnetic radiation.  $h$  is Planck's constant and  $\nu$  is the frequency of the emitted light.

In some chemiluminescent reactions the electronic energy of the excited-state molecule  $C^*$  is transferred to another molecule,

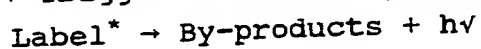
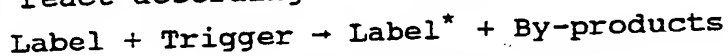
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which then decays to its ground-state by emitting a photon of electromagnetic radiation,



Specific binding assays, e.g. immunoassays, using chemiluminescent detection use one of the reactants as a label attached to one of the binding partners. In such assay, the reactants are generally called the label and the trigger and react according to the equation:



Examples of chemiluminescent labels which have been used in specific binding assays include acridinium esters, luminol, isoluminal, oxalate esters, dioxetanes, and luciferin. In many cases, the trigger molecule is an oxidant such as hydrogen peroxide which is capable of oxidizing the label in a highly energetic reaction which is capable of generating the excited state of the label.

Enhancer molecules are sometimes used in chemiluminescent reactions as a means of increasing the efficiency of the chemiluminescence process. Such molecules generally slow the reaction rate of the reaction and increase the quantum yield of the light emission.

Chemiluminescent binding assays have also been demonstrated in which an enzyme is used as the label. In these cases, the enzyme catalyzes the chemiluminescent reaction in the presence of a trigger solution. An example is the use of the enzyme horseradish peroxidase to catalyze the chemiluminescent reaction of luminol in the presence of hydrogen peroxide and hydroxide ion.

The term "chemiluminescent moiety," "label," "label compound," and "label substance," are used interchangeably. It is within the scope of the invention for the species termed "chemiluminescent moiety," "label compound," "label substance" and "label" to be linked to molecules such as an analyte or an analog thereof, a binding partner of the analyte or an analog thereof, and further binding partners of such aforementioned binding partner, or a reactive component capable of binding with the analyte, an analog thereof or a binding partner as mentioned above. The above-mentioned species can also be linked to a combination of one or more binding partners and/or one or more reactive

components. Additionally, the aforementioned species can also be linked to an analyte or its analog bound to a binding partner, a reactive component, or a combination of one or more binding partners and/or one or more reactive components. It is also within the scope of the invention for a plurality of the aforementioned species to be bound directly, or through other molecules as discussed above, to an analyte or its analog. For purposes of brevity, these ligands are referred to as an assay-performance-substance.

10           The terms detection and quantitation are referred to as "measurement", it being understood that quantitation may require preparation of reference compositions and calibrations.

15           The terms collection and concentration of complex may be used interchangeably to describe the concentration of complex within the assay composition and the collection of complex at, e.g., a surface of a flow cell.

20           Advantageously, the luminescence arises from chemiluminescence induced by exposing the label compound, whether bound or unbound to specific binding partners, to a trigger capable of triggering said label such that the label luminesces. Preferably, the trigger is an oxidant capable of oxidizing said label such that the label is oxidized and luminesces. The chemiluminescent reactive mixture is  
25           controllably triggered to emit light at a particular time and in a particular manner to generate light. Although the emission of visible light is an advantageous feature, the composition or system may emit other types of electromagnetic radiation, such as infrared or ultraviolet light,  
30           X-rays, microwaves, etc. Use of the terms "chemiluminescence" and "chemiluminescent" includes the emission of light and other forms of electromagnetic radiation.

### 35           Brief Description of the Drawings

Fig. 1 is a schematic drawing of a cell including a permanent magnet for performing the microparticulate-based nonseparation and separation assays of the invention.

Fig. 2 is a schematic representation of a direct incorporation PCR format using chemiluminescent labeled oligonucleotides and biotin chemiluminescent labeled oligonucleotides as primers.

5 Fig. 3 is a schematic representation of a normal PCR format using a biotinylated primer to allow the generation of biotinylated PCR and PRODUCT.

Fig. 4 is a schematic representation of an asymmetric PRC assay format generating single-stranded  
10 biotinylated DNA for later hybridization to chemiluminescent labeled oligonucleotides.

Fig. 5 is a schematic representation of a sedimentation assay cell which employs an electromagnet to cause the complex to settle in the collection zone surface.

15 Fig. 6 is a schematic representation of the lines of force in the vicinity of the collection zone as a function of the orientation of the magnet beneath the surface of the collection zone.

Fig. 7 is a schematic representation of a rotary  
20 flow cell wherein the complexes are deposited in the collection zone by centrifugation.

Fig. 8 is another schematic drawing of a cell including a permanent magnet for performing the microparticulate-based nonseparation and separation assays  
25 of the invention.

Fig. 9 is a simplified diagram of a voltage control apparatus for use with the cell of Fig. 8.

Fig. 10 is a schematic representation of an assay  
30 cell used to conduct assays relying upon gravitational force to cause the complex to settle.

#### Brief Description of the Invention

In its broadest embodiment, the invention is in a method for performing a binding assay for an analyte of  
35 interest present in a sample. The steps include:

- (a) forming a composition containing
  - (i) said sample;

- (ii) an assay-performance-substance which contains a component linked to a label compound capable of chemiluminescing when triggered; and
- 5 (iii) a plurality of particles capable of specifically binding with the analyte and/or said assay-performance-substance;
- (b) incubating said composition to form a complex which includes a particle and said label compound;
- 10 (c) collecting said complex in a collection zone;
- (d) introducing into said collection zone a trigger capable of triggering said label such that said label luminesces; and
- (e) measuring the emitted luminescence to measure the
- 15 presence of the analyte of interest in the sample.

Preferably, the trigger is an oxidant capable of oxidizing the label such that the label is oxidized and chemiluminescence. Also preferably, the particles are magnetically responsive and the complex is magnetically

20 collected in the collection zone.

In another embodiment, the assay-performance-substance or the particles further contain an enzyme for converting a precursor to an oxidant capable of oxidizing the label, and the trigger is the precursor.

25 While the invention is carried out by collecting the complex in a measurement zone, i.e., on a surface at which it can be caused to luminesce, the invention also embraces methods wherein the complex is collected in a measurement zone and thereafter means are brought to that

30 zone or other steps taken to induce and measure luminescence.

The collection of the complex may be carried out by several different methods, including gravity settling, filtration, centrifugation and magnetic attraction of

35 magnetically responsive particles which form part of the complex. The several embodiments are described in further detail below.

While batch assays can be performed, continuous or semi-continuous assays can be performed in flow cells. In a flow cell, the solid-phase remains in the measurement cell while the solution flows through the exits the cell. If the solid-phase (e.g., particles) are more dense than water, i.e., have a density greater than that of water, (more than 1.0 g/mL) the force of gravity upon the particles causes them to fall to the bottom of the cell. The cell can be constructed such that the particles settle to the bottom as the fluid flows through the cell or the cell can be constructed such that the majority of the sample is contained in the cell in a columnar compartment above the collection zone. Sufficient dwell time in the cell must be provided to permit the particles to settle in the collection zone before inducing chemiluminescence.

In another embodiment of the invention, the assay composition containing suspended particles having a density greater than the balance of the assay composition may be subjected to centrifugation in order to remove the particles to a measurement zone where they are subsequently brought into contact with, e.g., a trigger to induce chemiluminescence.

In this embodiment, the measurement cell is provided with means to rapidly rotate the sample and sample enclosure. Centrifugal force causes the particles in the sample to move outward from the axis of rotation of the sample enclosure and to collect on the outer surface of the sample enclosure.

In a third embodiment, the particles may be removed by filtration from the assay composition. In this embodiment the particles need not have a density greater than the balance of the assay composition. The invention, the particles are separated from the solution and concentrated by drawing the solution through a filter, e.g. pumping and collecting the particles on the surface of the filter.

In a preferred embodiment, the suspended particles are magnetically responsive, e.g. they may be paramagnetic

or ferromagnetic, and are collected in a measurement zone by imposition of a magnetic field on the particles. The measurement cell is equipped with a magnet. The magnetic field of the magnet applies a force on the particles as they reside in a batch cell or as they flow through a flow cell, causing them to separate from the bulk of the solution onto the surface of the cell which is in closest proximity to the magnet.

Several different heterogeneous and homogeneous formats for binding assays can be implemented using the methods described above to collect and concentrate the complex. In a heterogeneous binding assay the complex is separated from the composition before measuring luminescence from the label. In homogeneous assays, no separation of the bound (to the solid phase) and unbound labeled reagents is made.

In a homogeneous assay, when the complex is concentrated in a collection zone, the measured signal from the label is much greater than it would be in the absence of a concentration step. The signal from the uncomplexed labeled reagents, in contrast, is not changed. Hence, despite the presence of the uncomplexed labeled reagents in the measurement cell, the signal from the collected complex is stronger than in an assay without collection of complex. The detection limit for the binding assay is, much improved as a result of the collection procedure.

In a preferred embodiment of the invention, an *in-situ* separation step is included in the homogeneous binding assay procedure. After the assay composition, i.e., sample, assay performance substance and particles have been pumped into the measurement cell and the complex collected in a collection zone, a second fluid is pumped through the cell which is free of label or labeled reagents, thereby performing an *in-situ* wash or separation of the complex from unbound components of the assay composition. This assay procedure is technically a heterogeneous binding assay. However, the ability to perform the separation inside the measurement cell is advantageous in that it does not require

additional separation apparatus and the procedure is generally much faster than external separation methods.

Heterogeneous binding assays are conducted using the invention by mixing the components of the assay composition and allowing them to react for a predetermined length of time. The assay composition is then subjected to particles. Chemiluminescence is then measured from either the complex or the solution. Measuring the chemiluminescence from the complex after a concentration step permits measurement of analyte with better accuracy and with a lower detection limit than is possible without concentration.

In another embodiment, the invention is in a method for performing a binding assay for an analyte of interest present in a sample. The steps include:

- (a) forming a composition containing
  - (i) said sample;
  - (ii) an assay-performance-substance which contains a component linked to a label compound capable of chemiluminescing when triggered; and
  - (iii) a plurality of particles capable of specifically binding with the analyte and/or said assay-performance-substance;
- (b) incubating said composition to form a complex which includes a particle and said labeled component;
- (c) collecting said complex at the surface of an electrode;
- (d) inducing said label compound to luminesce by contacting it with a trigger, said trigger being formed in-situ by conversion of a precursor molecule upon introduction of electrochemical energy, for example by imposition of a voltage on said electrode; and
- (e) measuring the emitted luminescence to measure the presence of the analyte of interest in the sample.

The complex may be collected on, e.g., an electrode surface where it is excited and induced to chemiluminesce, as by introducing into a collection zone an

oxidant capable of oxidizing the label such that the label is oxidized and chemiluminescence. According to the present invention, the oxidant is formed in situ by conversion of a precursor molecule. The composition containing the sample  
5 may also contain the precursor molecule, or the precursor molecule may be introduced before incubating, after incubating but before magnetically collecting, or after magnetically collecting the incubated complex.

Thus, the invention relates to a method for  
10 performing a binding assay for an analyte of interest present in a sample including the steps of:

- (a) forming a composition containing
  - (i) said sample;
  - (ii) an assay-performance-substance which contains  
15 a component linked to a label compound capable of chemiluminescing when oxidized;
  - (iii) a plurality of coated magnetic particles capable of specifically binding with the analyte and/or said assay-performance-  
20 substance; and
  - (iv) a molecule capable of being electrochemically converted to an oxidant capable of oxidizing said label;
- (b) incubating said composition to form a complex  
25 which includes a particle and said label compound;
- (c) magnetically collecting said complex at the surface of an electrode;
- (d) converting said molecule to said oxidant by imposing a voltage on said electrode, such that said label  
30 is oxidized and luminesces; and
- (e) measuring the emitted luminescence to measure the presence of the analyte of interest in the sample.

The present invention further relates to a method for performing a binding assay for an analyte of interest  
35 present in a sample including the steps of:

- (a) forming a composition containing
  - (i) said sample;

- (ii) an assay-performance-substance which contains a component linked to a label compound capable of chemiluminescing when oxidized; and
- 5 (iii) a plurality of coating magnetic particles capable of specifically binding with the analyte and/or said assay-performance-substance;

10 (b) introducing to said composition a molecule capable of being electrochemically converted to an oxidant capable of oxidizing said label;

(c) incubating said composition to form a complex which includes a particle and said label compound;

15 (d) magnetically collecting said complex at the surface of an electrode;

(e) converting said molecule to said oxidant by imposing a voltage on said electrode, such that said label is oxidized and luminesces; and

20 (f) measuring the emitted luminescence to measure the presence of the analyte of interest in the sample.

The invention also relates to a method for performing a binding assay for an analyte of interest present in a sample including the steps of:

- (a) forming a composition containing
  - 25 (i) said sample;
  - (ii) an assay-performance-substance which contains a component linked to a label compound capable of chemiluminescing when oxidized; and
  - 30 (iii) a plurality of coated magnetic particles capable of specifically binding with the analyte and/or said assay-performance-substance;

35 (b) incubating said composition to form a complex which includes a particle and said label compound;

(c) introducing to said composition a molecule capable of being electrochemically converted to an oxidant capable of oxidizing said label;

(d) magnetically collecting said complex at the surface of an electrode;

(e) converting said molecule to said oxidant by imposing a voltage on said electrode, such that said label  
5 is oxidized and luminesces; and

(f) measuring the emitted luminescence to measure the presence of the analyte of interest in the sample.

The invention even further relates to a method for performing a binding assay for an analyte of interest  
10 present in a sample including the steps of:

(a) forming a composition containing

(i) said sample;

(ii) an assay-performance-substance which contains a component linked to a label compound  
15 capable of chemiluminescing when oxidized;  
and

(iii) a plurality of coating magnetic particles capable of specifically binding with the analyte and/or said assay-performance-  
20 substance;

(b) incubating said composition to form a complex which includes a particle and said label compound;

(c) magnetically collecting said complex at the surface of an electrode;

25 (d) introducing to said composition a molecule capable of being electrochemically converted to an oxidant capable of oxidizing said label;

(e) converting said molecule to said oxidant by imposing a voltage on said electrode, such that said label  
30 is oxidized and luminesces; and

(f) measuring the emitted luminescence to measure the presence of the analyte of interest in the sample.

In a preferred embodiment, the suspended particles are magnetically responsive, e.g. they may be paramagnetic  
35 or ferromagnetic, and are collected in a measurement zone or, preferably, directly at the surface of an electrode, by imposition of a magnetic field on the particles. The measurement cell is equipped with a magnet. The magnetic

field of the magnet applies a force on the particles as they reside in a batch cell or as they flow through a flow cell, causing them to separate from the bulk of the solution onto the surface of the cell which is in closest proximity to the magnet. If the magnet is placed in a proper orientation and in close proximity to the working electrode of a chemiluminescent detection system the particles will concentrate on the surface of the working electrode.

#### 10                    Detailed Description of the Invention

The invention, as well as other objects and features thereof, will be understood more clearly and fully from the following description of certain preferred embodiments.

15                    The invention is broadly applicable to analytes of interest which are capable of entering into binding reactions. These reactions include, e.g., antigen-antibody, ligand receptor, DNA and RNA interactions, and other known reactions. The invention relates to different methods and  
20 assays for qualitatively and quantitatively detecting the presence of such analytes of interest in a multicomponent sample.

#### The Samples

25                    The sample which may contain the analyte of interest, which may be in solid, emulsion, suspension, liquid, or gas form, may be derived from, for example, cells and cell-derived products, water, food, blood, serum, hair, sweat, urine, feces, tissue, saliva, oils, organic solvents  
30 or air. The sample may further comprise, for example, water, acetonitrile, dimethyl sulfoxide, dimethyl formamide, n-methyl-pyrrolidone or alcohols or mixtures thereof.

#### The Analytes

35                    Typical analytes of interest are a whole cell or surface antigen, subcellular particle, virus, prion, viroid, antibody, antigen, hapten, fatty acid, nucleic acid, protein, lipoprotein, polysaccharide, lipopolysaccharide,

glycoprotein, peptide, polypeptide, cellular metabolite, hormone, pharmacological agent, synthetic organic molecule, organometallic molecule, tranquilizer, barbiturate, alkaloid, steroid, vitamin, amino acid, sugar, lectin, 5 recombinant or derived protein, biotin, avidin, streptavidin, or inorganic molecule present in the sample. Typically, the analyte of interest is present at a concentration of  $10^{-3}$  molar or less, for example, as low as  $10^{-12}$  molar or lower.

10

#### Assay-Performance-Substance

The assay-performance-substance which is combined with the sample containing the analyte of interest contains at least one substance selected from the group consisting of 15 (i) added analyte of interest or its analog, as defined above, (ii) a binding partner of the analyte of interest or its said analog, and (iii) a reactive component, as defined above, capable of binding with (i) or (ii), wherein one of said substances is linked to a compound or moiety, e.g. a 20 chemiluminescent moiety capable of being induced to luminesce. The labeled substance may be a whole cell or surface antigen, a subcellular particle, virus, prion, viroid, antibody, antigen, hapten, lipid, fatty acid, nucleic acid, polysaccharide, protein, lipoprotein, 25 lipopolysaccharide, glycoprotein, peptide, polypeptide, cellular metabolite, hormone, pharmacological agent, tranquilizer, barbiturate, alkaloid, steroid, vitamin, amino acid, sugar, nonbiological polymer (preferably soluble), lectin, recombinant or derived protein, synthetic organic 30 molecule, organometallic molecule, inorganic molecule, biotin, avidin or streptavidin. In one embodiment, the reagent is an chemiluminescent moiety conjugated to an antibody, antigen, nucleic acid, hapten, small nucleotide sequence, oligomer, ligand, enzyme, or biotin, avidin, 35 streptavidin, Protein A, Protein G, or complexes thereof, or other secondary binding partner capable of binding to a primary binding partner through protein interactions.

5       Analog of the analyte of interest, which can be natural or synthetic, are typically compounds which have binding properties comparable to the analyte, but can also be compounds of higher or lower binding capability. The reactive components capable of binding with the analyte or its analog, and/or with a binding partner thereof, and through which the chemiluminescent moiety can be linked to the analyte, is suitably a second antibody or a protein such as Protein A or Protein G, or avidin or biotin or another component known in the art to enter into binding reactions.

10       The function of the chemiluminescent moieties is to emit electromagnetic radiation as a result of introduction into the reaction system of a trigger, particularly an oxidant. In order to do this, they must be capable of being stimulated to an excited energy state and also capable of emitting electromagnetic radiation, such as a photon of light, upon descending from that excited state.

20       The trigger is formed in-situ by conversion of a precursor molecule, for example water, to an oxidant, for example hydrogen peroxide or superoxide, upon introduction of electrochemical energy, for example by imposition of a voltage waveform on an electrode. As a second example, oxygen molecules as the precursor are electrochemically reduced to form hydrogen peroxide, the trigger.

25       The amount of chemiluminescent moiety incorporated in accordance with the invention will vary from system to system. Generally, the amount of such moiety utilized is that amount which is effective to result in the emission of a detectable, and if desired, quantifiable, emission of electromagnetic energy, from the aforementioned composition or system. The detection and/or quantitation of an analyte of interest is typically made from a comparison of the luminescence from a sample containing an analyte of interest and a chemiluminescent moiety to the luminescence emitted by a calibration standard developed with known amounts of the analyte of interest and chemiluminescent moiety. This assumes a homogeneous format. In the heterogeneous mode, a

separation as discussed previously is carried out prior to chemiluminescent analysis.

As can be appreciated by one of ordinary skill in the art, the identity and amount of the chemiluminescent moiety will vary from one system to another, depending upon prevailing conditions. The appropriate chemiluminescent moiety, and sufficient amount thereof to obtain the desired result, can be determined empirically by those of ordinary skill in the art, once equipped with the teachings herein, without undue experimentation.

#### The Particles

The particles advantageously comprise micro-particulate matter having a diameter of 0.05  $\mu\text{m}$  to 200  $\mu\text{m}$ , preferably 0.1  $\mu\text{m}$  to 100  $\mu\text{m}$ , most preferably 0.5  $\mu\text{m}$  to 10  $\mu\text{m}$ , and a surface component capable of binding to the analyte and/or one or more of the other substances defined in subparagraphs (b)(i), (b)(ii), or (b)(iii) above. For example, the microparticulate matter may be crosslinked starch, dextrans, cellulose, proteins, organic polymers, styrene copolymer such as styrene/butadiene copolymer, acrylonitrile/butadiene/styrene copolymer, vinylacetyl acrylate copolymer, or vinyl chloride/acrylate copolymer, inert inorganic particles, chromium dioxide, oxides of iron, silica, silica mixtures, and proteinaceous matter, or mixtures thereof. Desirably, the particles are suspended in the chemiluminescent system.

#### Apparatus for Measuring Electrochemiluminescence

An apparatus for carrying out the assays of the invention is described in Figs. 1. Fig. 1 discloses an advantageous chemiluminescent apparatus, but the methods of the present invention are not limited to application in apparatus 10, but rather may be employed in other types of chemiluminescent apparatus which include a means for collecting a labeled component. While the methods of the invention can be carried out in a static or flow-through mode, apparatus 10 includes a flow-through cell, which

provides distinct advantages for many types of samples including binding assay samples. Further details of apparatus for carrying out the chemiluminescent assays of the invention are disclosed in commonly assigned published PCT applications US 89/04854 and U.S. 90/01370.

Apparatus 10 includes a measurement cell 12, a light detection/measurement device 14, which may advantageously be a photomultiplier tube (PMT), photodiode, charge coupled device, photographic film or emulsion or the like, and a pump 16, which is advantageously a peristaltic pump, to provide for fluid transport to, through and from cell 12. Alternatively, a positive displacement pump may be used. A shutter mechanism 18 is provided between cell 12 and PMT 14 and is controllably operated to open only so far as to expose PMT 14 to cell 12 during chemiluminescent measurement periods. The shutter mechanism may be closed, for example, during maintenance. Also included in apparatus 10 but not illustrated in Fig. 1 is a lightproof housing intended to mount the various components therein and to shield PMT 14 from any external light during the chemiluminescent measurements.

Cell 12 itself includes a first mounting block 20 through which passes an inlet tube 22 and an outlet tube 24, which may be advantageously constructed of Plexiglas. Mounting block 20 has a first, outer surface 26 and a second, inner surface 28 defining one side of a sample-holding volume 30 of cell 12 in which cell 12 holds the cleaning and/or conditioning and/or measurement solutions during corresponding operations of apparatus 10. Inlet and outlet tubes 22, 24 pass through mounting block 20 from outer surface 26 to inner surface 28 and open into sample-holding volume 30. A second mounting block 32 is advantageously constructed of a material which is substantially transparent at the wavelength of chemiluminescent light emitted by the chemiluminescent moiety. Mounting block 32 is therefore advantageously formed of glass, plastic, quartz or the like and has a first, outer surface 34 and a second, inner surface 36.

Second mounting block 32 is separated from first mounting block 20 by an annular spacer 38, advantageously constructed of Teflon or other non-contaminable material. Thus, outer surface 34 of mounting block 30 defines the second side of the sample-holding volume 30. Spacer 38 has an outer portion 40 and a central aperture 42 whose inner edge 44 defines the side wall of sample-holding volume 30. Outer portion 40 seals the inner surface 28 of first mounting block 20 to outer surface 34 of second mounting block 32 to prevent any solution from passing out from sample-holding volume 30 between the two surfaces 28, 34.

Inlet tube 22 intersects sample-holding volume 30 at a first end 50 thereof adjacent to spacer 38 and outlet tube 24 intersects sample-holding volume 30 at a second end 52 thereof, adjacent spacer 38. The combination of inlet tube 22, sample-holding volume 30 and outlet tube 24 thereby provides a continuous flow path for the narrow, substantially laminar flow of a solution to, through and from cell 12.

Pump 16 is advantageously positioned at outlet tube 24 to "pull" solution from a sample volume in the direction of arrow A into inlet tube 22. The solution will flow through inlet tube 22, sample-holding volume 30 and outlet tube 24 and out in the direction of arrow B.

Alternatively, pump 16 may be positioned at inlet tube 22 to "push" the solution through apparatus 10. Advantageously, this same flow path through inlet tube 22, sample-holding volume 30 and outlet tube 24 is used for all solutions and fluids which pass through cell 12, whereby each fluid performs a hydrodynamic cleaning action in forcing the previous fluid out of cell 12. Pump 16 may be controlled to suspend its operation to hold a particular solution in cell 12 for any period of time.

Another apparatus for carrying out the assays of the invention is described in Figs. 8 and 9. Fig. 8 discloses an advantageous chemiluminescent apparatus, but the methods of the present invention are not limited to application in apparatus 10, but rather may be employed in

other types of chemiluminescent apparatus which include a working electrode or other triggering surface to provide electrochemical energy to convert the precursor to the trigger and a means for collecting a labeled component.

5 While the methods of the invention can be carried out in a static or flow-through mode, apparatus 10 includes a flow-through cell, which provides distinct advantages for many types of samples including binding assay samples. Further details of apparatus for carrying out the chemiluminescent  
10 assays of the invention are disclosed in commonly assigned published PCT applications US 89/04854 and U.S. 90/01370.

Apparatus 10 includes an electrochemical cell 12, a light detection/measurement device 14, which may advantageously be a photomultiplier tube (PMT), photodiode,  
15 charge coupled device, photographic film or emulsion or the like, and a pump 16, which is advantageously a peristaltic pump, to provide for fluid transport to, through and from cell 12. Alternatively, a positive displacement pump may be used. A shutter mechanism 18 is provided between cell 12  
20 and PMT 14 and is controllably operated to open only so far as to expose PMT 14 to cell 12 during chemiluminescent measurement periods. The shutter mechanism may be closed, for example, during maintenance. Also included in apparatus 10 but not illustrated in Fig. 1 is a lightproof housing  
25 intended to mount the various components therein and to shield PMT 14 from any external light during the chemiluminescent measurements.

Cell 12 itself includes a first mounting block 20 through which passes an inlet tube 22 and an outlet tube 24,  
30 which may be advantageously constructed of Plexiglas. Mounting block 20 has a first, outer surface 26 and a second, inner surface 28 defining one side of a sample-holding volume 30 of cell 12 in which cell 12 holds the cleaning and/or conditioning and/or measurement solutions  
35 during corresponding operations of apparatus 10. Inlet and outlet tubes 22, 24 pass through mounting block 20 from outer surface 26 to inner surface 28 and open into sample-holding volume 30. A second mounting block 32 is

advantageously constructed of a material which is substantially transparent at the wavelength of chemiluminescent light emitted by the chemiluminescent moiety. Mounting block 32 is therefore advantageously  
5 formed of glass, plastic, quartz or the like and has a first, outer surface 34 and a second, inner surface 36. Second mounting block 32 is separated from first mounting block 20 by an annular spacer 38, advantageously constructed of Teflon or other non-contaminable material. Thus, outer  
10 surface 34 of mounting block 30 defines the second side of the sample-holding volume 30. Spacer 38 has an outer portion 40 and a central aperture 42 whose inner edge 44 defines the side wall of sample-holding volume 30. Outer portion 40 seals the inner surface 28 of first mounting  
15 block 20 to outer surface 34 of second mounting block 32 to prevent any solution from passing out from sample-holding volume 30 between the two surfaces 28, 34.

Inlet tube 22 intersects sample-holding volume 30 at a first end 50 thereof adjacent to spacer 38 and outlet  
20 tube 24 intersects sample-holding volume 30 at a second end 52 thereof, adjacent spacer 38. The combination of inlet tube 22, sample-holding volume 30 and outlet tube 24 thereby provides a continuous flow path for the narrow, substantially laminar flow of a solution to, through and  
25 from cell 12.

Mounted on inner surface 28 of first mounting block 20 is a working electrode system 54 which, in the illustrated embodiment, includes working electrodes 58. In other embodiments, two or more working electrode may  
30 advantageously be provided. Working electrodes 58 is where the electrochemical and chemiluminescent reactions of interest can take place. Working electrodes 58 is a solid voltammetric electrode and may therefore be advantageously constructed of platinum, gold, carbons or other materials  
35 which are effective for this purpose. Wire connectors 62 connected to working electrodes 58 passes out through first mounting block 20.

Connector 62 is connected to a first, "working electrode" terminal 64 of a voltage control 66, illustrated in Fig. 2. Voltage control 66 advantageously operates in the manner of a potentiostat to supply voltage signals to working electrodes 58 and optionally to measure current flowing therefrom during an chemiluminescent measurement.

The potentiostat operation of voltage control 66 is further effected through a counter electrode 56 and, optionally but advantageously, a reference electrode 70. Counter electrode 56 and working electrodes 58 provide the interface to impress the potential on the solution within sample-holding volume 30 which energizes the chemical reactions and triggers electrochemical conversion of precursor to trigger in the sample and/or provides energy for cleaning and conditioning the surfaces of cell 12. Counter electrode 56 is connected by a wire connector 60 to a second, "counter electrode" terminal 78 of voltage control 66.

Reference electrode 70 provides a reference voltage to which the voltage applied by the working electrodes 58 is referred, for example, +1.2 volts versus the reference. Reference electrode 70 is advantageously located in outlet tube 24 at a position 80 spaced from cell 12 and is connected through a wire connector 82 to a third "reference electrode" terminal 84 of voltage control 66. In the three electrode mode, current does not flow through reference electrode 70. Reference electrode 70 may be used in a three electrode mode of operation to provide a poised, known and stable voltage and is therefore advantageously constructed of silver/silver chloride (Ag/AgCl) or is a saturated calomel electrode (SCE). Voltage control 66 may be operable in a two electrode mode of operation using only working electrode 58 and electrode 56 as a counter/reference electrode. In this two electrode mode of operation, counter/reference electrode 56 is electrically connected to voltage control terminals 78 and 84 on voltage control 66. In this case, voltage control 66 operates essentially as a battery. Voltage control 66 supplies voltage signals to

working and counter electrodes 58 and 56 and optionally measures the current flowing through the respective electrodes. Reference electrode 70 may alternatively be a so-called "quasi-reference" electrode constructed of

5 platinum, gold, stainless steel or other material, which provides a less stable voltage, yet one that is measurable with respect to the solution in contact. In both the two and three electrode mode, the reference electrode 70 or 56 serves the purpose of providing a reference against which

10 the voltage applied to working electrodes 58 is measured. The poised voltage reference is currently considered to be more advantageous. Voltage control 66 in its potentiostat operation controls the various electrodes by providing a known voltage at working electrodes 58 with respect to

15 reference electrode 70 while measuring the current flow between working electrodes 58 and counter electrode 56. Potentiostats for this purpose are well known, and the internal structure of voltage control 66 may therefore correspond to any of the conventional, commercially avail-

20 able potentiostats which produce the above-recited functions and so do not form a part of the present invention per se. Indeed, apparatus 10 may alternatively be constructed without an internal voltage control 66, and may be adapted to be connected to an external potentiostat which is

25 separately controlled for providing the required voltage signals to electrodes 56, 58, 72, 74 and 70. These voltage signals, applied in a specific manner as described below, provide repeatable initial conditions for the surfaces of working electrodes 58 and advantageously for the surfaces of

30 cell 12 as a whole, a feature which contributes significantly to improved precision in chemiluminescent measurements.

Pump 16 is advantageously positioned at outlet tube 24 to "pull" solution from a sample volume in the

35 direction of arrow A into inlet tube 22. The solution will flow through inlet tube 22, sample-holding volume 30 and outlet tube 24 past reference electrode 70 and out in the direction of arrow B. Alternatively, pump 16 may be

positioned at inlet tube 22 to "push" the solution through apparatus 10. Advantageously, this same flow path through inlet tube 22, sample-holding volume 30 and outlet tube 24 is used for all solutions and fluids which pass through cell 12, whereby each fluid performs a hydrodynamic cleaning action in forcing the previous fluid out of cell 12. Pump 16 may be controlled to suspend its operation to hold a particular solution in cell 12 for any period of time.

The flow-through construction of apparatus 10 permits working electrodes to be impressed with a variable voltage or to be continuously held at a preoperative potential while being continuously exposed to one or more solutions without exposing working electrodes 58 (or counter and reference electrodes 56, 70) to air. Exposure to air, which opens the circuit to the reference electrode 70, permits unknown, random voltage fluctuations which destroy the reproducibility of surface conditions on working electrodes 58. The flow-through construction permits the rapid alternation between initializing steps, in which electrode system 54 is cleaned and conditioned, and measurement steps, in which one or more measurement waveforms or sweeps trigger chemiluminescent.

#### Assay Media

In order to operate a system in which an electrode introduces electrochemical energy to convert the precursor molecule to the oxidant such that said label is oxidized and luminesces, it is necessary to provide an electrolyte in which the electrode is immersed and which contains the precursor molecule. The electrolyte is a phase through which charge is carried by ions. Generally, the electrolyte is in the liquid phase, and is a solution of one or more salts or other species in water, an organic liquid or mixture of organic liquids, or a mixture of water and one or more organic liquids. However, other forms of electrolyte are also useful in certain embodiments of the invention. For example, the electrolyte may be a dispersion of one or more substances in a fluid --e.g., a liquid, a vapor, or a

supercritical fluid --or may be a solution of one or more substances in a solid, a vapor or supercritical fluid.

The electrolyte is suitably a solution of a salt in water. The salt can be a sodium salt or a potassium salt preferably, but incorporation of other cations is also suitable in certain embodiments, as long as the cation does not interfere with the chemiluminescent interaction sequence. The salt's anion may be a phosphate, for example, but the use of other anions is also permissible in certain embodiments of the invention --once again, as long as the selected anion does not interfere with the chemiluminescent interaction sequence.

The composition may also be nonaqueous. While supercritical fluids can in certain instances be employed advantageously, it is more typical to utilize an electrolyte comprising an organic liquid in a nonaqueous composition. Like an aqueous electrolyte, the nonaqueous electrolyte is also a phase through which charge is carried by ions. Normally, this means that a salt is dissolved in the organic liquid medium. Examples of suitable organic liquids are acetonitrile, dimethylsulfoxide (DMSO), dimethylformamide (DMF), methanol, ethanol, and mixtures of two or more of the foregoing. Illustratively, tetraalkylammonium salts, such as tetrabutylammonium tetrafluoroborate, which are soluble in organic liquids can be used with them to form nonaqueous electrolytes.

The electrolyte is, in certain embodiments of the invention, a buffered system. Phosphate buffers are often advantageous. Examples are an aqueous solution of sodium phosphate/sodium chloride, and an aqueous solution of sodium phosphate/sodium fluoride.

#### Other Aspects of the Invention

The invention is also directed to reagent compositions. Broadly, the reagent compositions may be any one of the components of the assay systems of the invention, i.e., (a) electrolyte, (b) label compound containing an chemiluminescent moiety, (c) particles, including

magnetically responsive particles, (d) analyte of interest or an analog of the analyte of interest, (e) a binding partner of the analyte of interest or of its analog, (f) a reactive component capable of reacting with (d) or (e), (g) a trigger precursor molecule, or (h) a chemiluminescence-reaction enhancer. The reagents may be combined with one another for convenience of use, i.e., two component, three component, and higher multiple component mixtures may be prepared, provided that the components are not reactive with one another during storage so as to impair their function in the intended assay. Desirably, the reagents are two-component or multicomponent mixtures which contain particles as well as one or more other components.

The invention is also directed to kits. The kits may include vessels containing one or more of the components (a) to (h) recited above or the kits may contain vessels containing one or more reagent compositions as described above comprising mixtures of those components, all for use in the assay methods and systems of the invention.

20

#### Description of Preferred Embodiments of the Invention

While a wide range of particles can be employed in the particle-based assays of the invention, generally the particles have a density of from 1.0 to 5.0 g/mL and preferably have a density of from 1.1 to 2 g/mL. Choice of the optimum density is within the skill of the art, the rate of settling in gravity-driven assays being a trade-off between the speed of the assay and the desire to create a uniform layer of complex in the collection zone.

Particles having a wide range of mean diameters can also be employed. Particles having a mean diameter of from 0.001 to 100  $\mu\text{m}$  can be used and preferably the particles have a mean diameter of from 0.01 to 10  $\mu\text{m}$ .

Wide ranges of concentration of particles in the assay composition can also be employed. For example, the concentration can range from 1 to 10,000  $\mu\text{g/mL}$  to preferably from 5 to 1000  $\mu\text{g/mL}$ . Desirably, the density of the particles, their size and their concentration is selected

such that the particles settle at a rate of at least 0.5 mm/min and preferably at a faster rate.

In the filtration mode of performing the invention, the filtration means desirably has a pore size, measured as mean diameter, from broadly 0.01 to 90% of the mean diameter of the particles and preferably from 10% to 90% of that diameter.

The art has described a number of magnetic particles which can be used in the assays of the invention. For example, U.S. Patent No. 4,628,037, 4,695,392, 4,695,393, 4,698,302, 4,554,088, U.K. Patent Application GB 2,005,019A and EP 0,180,384, describe a variety of magnetic particles which can be used with success. The particles may be paramagnetic or ferromagnetic and may be coated with various materials to which binding compounds are coupled so that the magnetic particle can be used in immunoassays. Desirably the magnetic particles used in the invention have a susceptibility of at least 0.001 cgs units and desirably the susceptibility is at least 0.01 cgs units. The magnetic particles may have a broad range of densities, i.e. from substantially less than that of water, 0.01, to 5 g/mL and preferably from 0.5 to 2 g/mL. The particle sizes can range from 0.001 to 100  $\mu$ m and preferably from 0.01 to 10  $\mu$ m. The concentration of the particles may range broadly from 1 to 10,000  $\mu$ g per mL and preferably is from 5 to 1000  $\mu$ g per mL.

Desirably the magnetic particles which are used have a low magnetic remanence, as described for example EP 0,180,384, so that after the magnetic field is removed from the electrode surface, the particles demagnetize and can be swept out of the assay cell. Desirably the density, concentration and particle size of the magnetic particles is chosen such that the settling time is at least 0.5 mm/min and desirably it is above that rate.

### 35 Assays

A variety of assays can be performed using the methods of the invention.

An assay was performed as shown in Fig. 2. The products resulting from the reaction are labeled with biotin and a chemiluminescent label. Streptavidin beads capture the bifunctionalized DNA via biotin streptavidin binding and this is followed by washing. The bead bound product is then subjected to analysis detecting the chemiluminescent label.

An assay was performed as shown in Fig. 3. The biotinylated PCR product was captured on streptavidin beads and the non-biotinylated strand removed. The bead bound PCR product is then hybridized with a chemiluminescent labeled oligonucleotide. This is followed by chemiluminescent analysis to detect the label.

An assay was conducted as shown in Fig. 4. The hybrids were captured on streptavidin beads. This is followed by chemiluminescent analysis without washing.

#### EXAMPLES 1 - 11

##### Instrumentation, Materials, and Methods

###### (1) Instrumentation

A flow-through apparatus as described in Fig. 1 was used.

Teflon Gasket (0.15" thick)

Plexiglas Faceplate

Inlet Tubing = .042" id polypropylene

Aspiration Rates: variable from 0.01 to 5 mL/min

Luminometer using Hamamatsu R374 PMT (low gain red sensitive tube); PMT Voltage variable 0-1400 V

#### EXAMPLE 1

##### Chemiluminescent Apparatus and Method for Deposition of Microparticles

Magnetic Collection using a Sedimentation Cell.

A cell for conduct of an assay using magnetic force to cause the microparticulate to settle is shown in Fig. 5. Reference numeral 21 refers to a transparent window, reference numeral 22 to a gasket, reference numeral 23 to the inlet in the cell block, reference numeral 25 to

the sample outlet, reference numeral 26 to the cell block itself and reference 27 to an electromagnet.

The plane of the cell block is oriented horizontally. Labeled microparticles (Dynal) in buffer are drawn to the cell by means of a peristaltic pump. The pump is turned off after the microparticles reach the cell. The microparticles in the cell chamber are drawn to the collection zone by means of a magnetic field generated using electromagnet 27 operating at 12 volts and 1.5 amps. By application of the electromagnet, the rate of deposition of microparticles is greatly increased over that observed when the microparticles settle solely due to the force of gravity.

15

#### EXAMPLE 2

##### Chemiluminescent Apparatus and Method for Deposition of Microparticles

##### Magnetic Collection using a Collection Cell.

An assay is carried out in a cell as described in Fig. 1. With reference to Fig. 1, reference numeral 32 refers to transparent window, reference numeral 38 to a gasket, reference numeral 22 to an inlet in the cell block, reference numeral 20 to the cell block itself, reference numeral 24 to the sample outlet and reference numeral 37 to a permanent magnet.

The plane of the cell block is oriented horizontally. Labeled microparticles (Dynal) in buffer are drawn to the cell by means of a peristaltic pump 11. Prior to the sample introduction, permanent magnet 37 is positioned immediately below the collection zone at a distance of 0.035 inches. As the sample is being drawn to the cell, the microparticles collect in a collection zone, as defined by the area of the magnet. The pump is turned off. The longer the collection time, the more particles are collected.

EXAMPLE 3Use of Magnet for Deposition of Microparticles  
Magnetic Field Orientation.

5 Microparticles which are attracted to a magnet  
whether a permanent magnet or electromagnet, align with the  
orientation of the magnetic field. Fig. 6 depicts magnetic  
fields and the resultant particle arrangements which are  
parallel (A) and perpendicular (B) to the top surface of  
cell blocks 8 and 4, respectively, in the vicinity of that  
10 surface. One skilled in the art will appreciate that the  
orientation of the particles in the collection zone will  
affect the efficiency of subsequent contact with trigger.

EXAMPLE 4Particle Collection and Concentration by Filtration

15 Microparticles which are magnetically responsive,  
non-magnetically responsive, and of a wide range of  
densities can advantageously be collected by filtration upon  
the surface of a membrane filter. In one embodiment of the  
20 invention, the particles are pumped through a portion of a  
filter membrane which has pore sizes which are smaller than  
the diameter of the particles but preferably are  
substantially smaller than the particle diameter and at a  
sufficiently high surface density such that the collection  
25 of particles will not cause blockage of the pores. The  
collected particles are then exposed to trigger by pumping  
the trigger solution through the filter for the purpose of  
inducing chemiluminescence from the particles and measuring  
the luminescence to measure the quantity of chemiluminescent  
30 label on the particles.

In another embodiment, the membrane filter having  
pore sizes as described above is attached or placed upon the  
surface of an absorbent material such that capillarity or  
"wicking" will spontaneously draw fluids containing  
35 microparticles through the membrane filter without requiring  
any apparatus to induce the flow of fluid through the  
filter.

In a preferred embodiment, the membrane filter, having pore sizes as described above, is coated with a thin film of metal or other electronically conductive material such that the surface of the membrane can serve as a working  
5 electrode in the chemiluminescent apparatus. The conductive films are readily applied to the surface of a membrane by methods commonly used in the fabrication of microelectronic devices, e.g., thermal evaporation or sputtering.

Such a filter is readily mounted in a flow cell  
10 such that the flow-path for the fluid is through the filter. Particles in the stream are trapped by the filter and are easily washed *in-situ* providing for a rapid and simple means for performing heterogeneous assays without any external washing apparatus.

15

#### EXAMPLE 5

##### Particle Collection and Concentration by Centrifugal Method

The rotary flow cell shown in Fig. 7 provides  
20 another means to collect the complex in order to measure luminescence. The assay solution 61 is pumped into cell 62 through rotary seal 63 while a rotational motion is imparted to the cell. The denser particles of the complex are concentrated in the collection zone. While the cell is  
25 still rotating the solution passes out of the cell. The light output passing through cell window 67 is measured by photomultiplier tube 65. The light output is directed from the collection zone and reflecting off curved mirror surface 66 located at the center of the cell. The cell is then  
30 flushed and cleaned for the next cycle. This may be accomplished with the cell stopped or rotating.

#### EXAMPLE 6

##### Coating of Particles With Labeled Non-specific Protein 35 at Moderate Surface Concentration

30 mg (1 ml) of 4.5 um uncoated magnetically responsive, polystyrene M-450 DYNABEADS (DYNAL, Oslo, Norway) were washed by magnetic separation with a 150 mM

phosphate buffer pH 7.5 solution using 2 ml/wash. 150  $\mu$ g of acridinium ester-labeled antibody (London Diagnostics LumaTag TSH Labeled Antibody) in 1 ml of phosphate buffer saline (PBS) with 0.05% thimerasol is added to the particles. This mixture is allowed to incubate overnight at room temperature with agitation. The solution was then magnetically separated from the particles and the fluid removed. To block unreacted sites, 1 ml of 3% BSA/PBS with 0.05% sodium azide was added to the particles, and the resultant solution was allowed to incubate 2 hours at room temperature. The particles were washed 5 times (2 ml/wash), and then finally resuspended in 6 ml of the same buffer for storage.

#### EXAMPLE 7

##### Chemiluminescent Measurement Using Magnetically Responsive Particles

Magnetically responsive particles (Dynal, Oslo, Norway) are coated with labeled proteins as described in Example 6. The coated particles are washed with phosphate buffer three times before making 2 mL of a 30  $\mu$ g/mL suspension in 0.1 N  $\text{HNO}_3$  and 0.5% hydrogen peroxide. Using a peristaltic pump, 500  $\mu$ l of the particle suspension is drawn into the flow cell (Example 2). As the particles flow through the cell, they are attracted and concentrated into the collection zone by a magnet. After the particles are magnetically collected, a solution of 0.25 N NaOH, 0.5% hydrogen peroxide is drawn through the cell while the chemiluminescence is measured using a Hamamatsu R374 photomultiplier tube centered above the flow cell where particles have concentrated in the collection zone.

#### EXAMPLE 8

##### Preparation Of Physically Adsorbed Sheep Anti-Thyroid Stimulating Hormone (TSH) Coated Dynal Particles

1 mL of 4.5 $\mu$ m uncoated magnetic, polystyrene particles with -OH residues on their surfaces (DYNAL, DYNABEADS M-450, DYNAL A.S. Oslom Norway) is washed by

magnetic separation with a 150mM sodium carbonate/bicarbonate pH 9.6 solution using 2mL/wash. 0.5mg of purified monoclonal anti-TSH antibody (Catalog No. 5064031, Ventrex Laboratories, Inc., Portland, Maine) in 1 mL. of the carb/bicarb solution is added to the particles. This mixture is incubated overnight at room temperature with mixing. The solution is then magnetically separated from the particles and removed. 1mL of 3% BSA/PBS with 0.05% sodium azide is added and incubated 2 hours at room temperature with agitation to block unreacted sites. The particles are washed 5 times (2mL/wash) and then finally resuspended in 1mL of the same buffer for storage. The final concentration is 3% by weight.

15

EXAMPLE 9One Step Separation Sandwich Assay for Thyroid Stimulating Hormone (TSH)

100  $\mu$ L serum calibrators (London Diagnostics TSH LumiTAG Kit), 25 $\mu$ L LumaTag TSH acridinium ester-labeled antibody (London Diagnostics) in phosphate buffer and 25  $\mu$ L anti-TSH-DYNAL particles (Example 8) in phosphate buffer are combined and incubated in polypropylene tubes for 15 minutes, at room temperature, with mixing. The particles are then washed by magnetic separation and then resuspended in 500 $\mu$ L of pH 4, 10mM carbonate/bicarbonate buffer. This wash procedure was repeated two additional times. The particles are drawn into a flow cell (Example 2), magnetically collected and a solution to trigger the chemiluminescent reaction is drawn through the flow cell (0.5% hydrogen peroxide, 0.25 N NaOH). The chemiluminescence for each sample is measured as described in Example 2. The chemiluminescent intensity is directly proportional to the concentration of analyte present in the sample (increasing intensity as the concentration of analyte increases).

EXAMPLE 10One Step Non Separation Sandwich Assay for Thyroid  
Stimulating Hormone (TSH)

100 $\mu$ L serum calibrators (London Diagnostics TSH  
5 LumiTAG Kit), 25 $\mu$ L LumaTag TSH acridinium ester-labeled  
antibody (London Diagnostics) in phosphate buffer and 25 $\mu$ L  
anti-TSH-DYNAL particles (Example \*) in phosphate buffer are  
combined and incubated in polypropylene tubes for 15  
minutes, at room temperature, with mixing. Prior to reading  
10 results, 1mL of pH 4 100 mM carbonate/bicarbonate buffer is  
added. The particles are drawn into a flow cell (Example  
2), magnetically collected and a solution to trigger the  
chemiluminescent reaction is drawn through the flow cell  
(0.5 % hydrogen peroxide, 0.25 N NaOH). The  
15 chemiluminescence for each sample is read as described in  
Example 2. The chemiluminescent intensity is directly  
proportional to the concentration of analyte present in the  
sample (increasing intensity as the concentration of analyte  
increases).

20

EXAMPLE 11Chemiluminescent TSH Immunoassay Using  
Enzyme to Generate Trigger

Using magnetically responsive microparticles as  
25 the solid phase and acridinium ester as the label, a TSH  
immunoassay can be performed using the apparatus described  
in Example 2. An enzyme, glucose oxidase is used to convert  
a precursor of the trigger (glucose) to the trigger  
(hydrogen peroxide). The enzyme glucose oxidase catalyzes  
30 the oxidation of glucose to gluconic acid and hydrogen  
peroxide. Acridinium ester in the presence of hydrogen  
peroxide is oxidized to an excited state. The subsequent  
return to ground state of oxidized excited product results  
in the emission of light which is quantified. Magnetic  
35 microparticles coated with both specific antibody and enzyme  
glucose oxidase can be used (prepared as in Example 8 except  
that 0.5 mg of both antibody and enzyme (Sigma Chemical) are  
added to the particles for coating). Alternatively,

separate particles coated with each reagent (antibody or enzyme) can be mixed and used in the assay (prepared separately as described in Example 8).

- 5 D-Glucose Glucose Oxidase > D- gluconic acid + H<sub>2</sub>O<sub>2</sub>  
Acridinium Ester + H<sub>2</sub>O<sub>2</sub> -----> Light (428 nm)

The TSH immunoassay is based on a two-site sandwich assay known in the art. Monoclonal anti-TSH antibody coated magnetic microparticles are prepared as described in Example 8. Enzyme glucose oxidase coated magnetic microparticles are prepared by the same method as antibody coated magnetic microparticles. Acridinium Ester labeled polyclonal anti-TSH antibody and the TSH standards are obtained from London Diagnostics. Enzyme substrate solution consists of 100 mM potassium phosphate buffer containing D-glucose (100 mg/ml).

A series of tubes (12 x 75 mm polypropylene) are set up and labeled according to standards and samples to be assayed. Into each tube is added 100  $\mu$ l of standard or sample or control, 100  $\mu$ l of acridinium ester-labeled antibody and 100  $\mu$ l of a mixture of anti-TSH antibody and enzyme coated microparticles. The tubes are incubated at room temperature with mixing for 15 min. Following incubation, 1 ml pH 4, 100 mM carbonate/bicarbonate buffer is added. The particles are drawn into a flow cell (Example 2), magnetically collected and the glucose substrate solution is drawn through the flow cell. The chemiluminescence for each sample is read as described in Example 2. The chemiluminescent intensity is directly proportional to the concentration of analyte present in the sample (increasing intensity as the concentration of analyte increases).

#### EXAMPLES 12 - 14

#### 35 Instrumentation, Materials, and Methods

##### (1) Instrumentation

A flow-through apparatus, employing three electrodes, as described in Figs. 8 and 9, was used.

40

Working Electrode -- Au disk, 3 mm diameter  
Counter Electrode -- Au disk, 3 mm diameter  
Reference Electrode -- Ag/AgCl  
Teflon Gasket (0.15"thick)  
5 Plexiglas Faceplate  
Inlet Tubing = .042" id polypropylene  
Aspiration Rates:variable from 0.01 to 5 mL/min  
Potentiostat: microprocessor controlled  
Luminometer using Hamamatsu R374 PMT (low gain red  
10 sensitive tube); PMT Voltage variable 0-1400 V

#### EXAMPLE 12

##### Apparatus and Method for Collection of Microparticles by Gravity

15 The measurement is conducted in a cell as shown in Fig. 10. References made to Fig. 10 which depict an apparatus for conducting an assay using the force of gravity. The components of the apparatus include a transparent window identified by reference numeral 11, a  
20 gasket identified by reference numeral 12, a block which includes an inlet 13, a working electrode 14, a counterelectrode 15 and an outlet port 16. The plane of the cell block is horizontal, i.e. perpendicular to the direction of the earth's gravitational field. Labeled  
25 microparticles (Dynal) in an buffer are drawn to the cell by means of a peristaltic pump. The pump is turned off after the particles reach the cell. The microparticles in the cell chamber fall onto the working electrode surface. The rate of fall of microparticles is determined to be  
30 approximately constant at 0.5 mm/min over a distance of 10 mm. The number of particles to settle is a function of time and rate of fall. The chemiluminescent intensity is proportional to the number of particles that settle on the working electrode. The number of particles that reach the  
35 surface, and therefore the chemiluminescent intensity, is limited by the height of fluid sample over the working electrode.

EXAMPLE 13Chemiluminescence Apparatus and Method for Deposition of  
MicroparticlesMagnetic Collection using a Collection Cell

5           An assay is carried out in a cell as described in  
Fig. 8. With reference to Fig. 8, reference numeral 32  
refers to transparent window, reference numeral 38 to a  
gasket, reference numeral 22 to an inlet in the cell block,  
reference numeral 58 to a working electrode, reference  
10 numeral 35 to the cell block itself, reference numeral 24 to  
the sample outlet and reference numeral 37 to a permanent  
magnet.

          The plane of the cell block is oriented  
horizontally. Labeled microparticles (Dyna) in  
15 chemiluminescent buffer are drawn to the electrochemical  
cell by means of a peristaltic pump. Prior to the sample  
introduction, permanent magnet 34 is positioned immediately  
below the working electrode/solution interface at a distance  
of 0.035 inches. As the sample is being drawn to the cell,  
20 the microparticles collect in a collection zone, for example  
deposit onto an area over the working electrode, as defined  
by the area of the magnet. The pump is turned off and the  
magnet withdrawn after the entire sample is deposited. The  
longer the collection time, the more particles are  
25 deposited. Increasing the concentration of particles on the  
working electrode results in an increased chemiluminescent  
intensity.

EXAMPLE 14

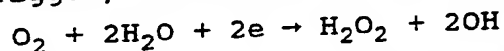
30           Chemiluminescent Measurement Using Magnetically  
Responsive Particles

          Magnetically responsive particles (Dyna, Oslo,  
Norway) are coated with labeled proteins as described in  
Example 6. The coated particles are washed with phosphate  
35 buffer three times before making 2 mL of a 30 ug/mL  
suspension in pH 4, 100mM carbonate/bicarbonate buffer.  
Using a peristaltic pump, 500 ul of the particle suspension  
is drawn into the flow cell. As the particles flow to the

working electrode, they are attracted and concentrated onto the working electrode by a magnet. After the particles are magnetically collected, a solution of 0.25 N NaOH is drawn through the cell. The chemiluminescence is generated by conversion of precursor molecules to trigger molecules by one of three methods:

(1) a potential waveform is applied to the electrochemical electrodes such that water (precursor) is oxidized, forming hydrogen peroxide and other oxidizing species (triggers) at the working electrode. The hydrogen peroxide or other oxidizing species triggers the chemiluminescent reaction of the acridinium ester label.

(2) a potential waveform is applied to the electrochemical electrodes such that oxygen (precursor) dissolved in the solution is reduced to form hydrogen peroxide (trigger) according to the equation



This electrochemical reaction occurs upon application of approximately -0.4 V to the working electrode relative to a saturated calomel electrode. The hydrogen peroxide triggers the chemiluminescent reaction of the acridinium ester label.

(3) a potential waveform is applied to the electrochemical electrodes such that oxygen is produced at the working electrode by oxidative electrolysis of water. A waveform is then applied to the electrochemical electrodes such that oxygen (precursor) created by the electrolysis of water is reduced to form hydrogen peroxide (trigger). The hydrogen peroxide triggers the chemiluminescent reaction of the acridinium ester label.

The chemiluminescence is measured using a Hamamatsu R374 photomultiplier tube centered above the flow cell where particles have concentrated in the collection zone on the working electrode.

**WHAT IS CLAIMED:**

1. A method for performing a binding assay for an analyte of interest present in a sample comprising the steps of:
  - 5 (a) forming a composition containing
    - (i) said sample;
    - (ii) an assay-performance-substance which contains a component linked to a label compound capable of chemiluminescing when triggered;
    - 10 and
    - (iii) a plurality of particles capable of specifically binding with the analyte and/or said assay-performance-substance;
  - (b) incubating said composition to form a complex
  - 15 which includes a particles and said labeled component;
  - (c) collecting said complex in a collection zone;
  - (d) introducing into said collection zone a trigger capable of triggering said label such that said label luminesces; and
  - 20 (e) measuring the emitted luminescence to measure the presence of the analyte of interest in the sample.
2. A method as recited in claim 1 wherein said trigger is an oxidant capable of oxidizing capable said label.
- 25 3. A method as recited in claim 1 wherein said particles are magnetically responsive and said complex is magnetically collected in said collection zone.
4. A method as recited in claim 2 wherein said oxidant is hydrogen peroxide or superoxide.
- 30 5. A method as recited in claim 1 wherein assay-performance-substance further contains an enzyme for converting a precursor to an oxidant capable of oxidizing said label, and said trigger is the precursor.
6. A method as recited in claim 5 wherein the
- 35 enzyme is glucose oxidase, the precursor is glucose and the oxidant is hydrogen peroxide.
7. A method as recited on claim 1 wherein said particles further contain an enzyme for converting a

precursor to an oxidant capable of oxidizing said label, and said trigger is the precursor.

8. A method as recited in claim 7 wherein the enzyme is glucose oxidase, the precursor is glucose and the oxidant is hydrogen peroxide.

9. A method as recited in claim 1 conducted as a batch process, the composition being permitted to reside within said cell for a time sufficient to permit collection of said particles.

10. A method as recited in claim 1 conducted as a flow process wherein said composition is flowed through said cell at a sufficiently low rate to permit collection of at least a portion of said particles.

11. An assay method as recited in claim 7 wherein said particles have a density of from 0.1 to 5g/mL.

12. An assay method as recited in claim 11 wherein said particles have a density of from 0.5 to 2g/mL.

13. An assay method as recited in claim 7 wherein the size of said particles, measured as the mean diameter, ranges from 0.001 to 100  $\mu\text{m}$ .

14. An assay method as recited in claim 13 wherein said the size of said particles range from 0.01 to 10  $\mu\text{m}$ .

15. An assay method as recited in claim 7 wherein the concentration of particles in said composition is from 1 to 10,000  $\mu\text{g/mL}$ .

16. An assay method as recited in claim 15 wherein said concentration of particles is in the range of from 5 to 1000  $\mu\text{g/mL}$ .

17. An assay method as recited in claim 7 wherein said particles have a magnetic susceptibility of at least 0.001 cgs units.

18. A method as recited in claim 17 wherein the magnetic susceptibility is at least 0.01 cgs units.

19. An assay method as recited in claim 7 wherein the magnetic susceptibility, density, size and concentration of said particles in said composition is such that the settling rate of said particles is at least 0.5 mm/min.

20. An apparatus for performing a binding assay for an analyte of interest present in a sample based upon measurement of chemiluminescence comprising:

(a) a cell defining a sample containing volume having  
5 a vertical columnar zone and having inlet and outlet means, and further including means for generating a magnetic field positioned below a substantial volume of said cell and said columnar zone; and

(b) means to measure the chemiluminescence generated  
10 at a collection zone.

21. An apparatus as recited in claim 20 wherein said means for generating a magnetic field includes a plurality of magnets in north-south orientation and separated by non-magnetic material.

15 22. A method for performing a binding assay for an analyte of interest in a sample comprising the steps of:

(a) forming a composition containing

(i) said sample;

(ii) an assay-performance-substance which contains  
20 a component linked to a label compound capable of chemiluminescing when triggered; and

(iii) a plurality of particles capable of specifically binding with the analyte and/or  
25 said assay-performance-substance;

(b) incubating said composition to form a complex which includes a particles and said label component;

(c) collecting said complex at the surface of an electrode;

30 (d) inducing said label to luminesce by contacting it with a trigger, said trigger being formed *in-situ* by conversion of a precursor molecule upon introduction of electrochemical energy; and

(e) measuring the emitted luminescence to measure the  
35 presence of the analyte of interest in the sample.

23. A method as recited in a claim 22 wherein said trigger is an oxidant capable of oxidizing said label.

24. A method as recited in claim 22 wherein said particles are magnetically responsive and said complex is magnetically collected at the surface of the electrode.

5 25. A method as recited in claim 22 wherein the magnetic field employed to collect said complex at the surface of said electrode is substantially dissipated before measuring the emitted luminescence.

26. A method as recited in claim 22 wherein said molecule is water, and said oxidant is hydrogen peroxide or  
10 superoxide.

27. A method as recited in claim 22 wherein said molecule is oxygen, and said oxidant is hydrogen peroxide.

28. A method for performing a binding assay for an analyte of interest present in a sample comprising the  
15 steps of:

(a) forming a composition containing

(i) said sample;

20 (ii) an assay-performance-substance which contains a component linked to a label compound capable of chemiluminescing when oxidized;

(iii) a plurality of coated magnetic particles capable of specifically binding with the analyte and/or said assay-performance-substance; and

25 (iv) a molecule capable of being electrochemically converted to an oxidant capable of oxidizing said label;

(b) incubating said composition to form a complex which includes a particle and said label compound;

30 (c) magnetically collecting said complex at the surface of an electrode;

(d) converting said molecule to said oxidant by imposing a voltage on said electrode, such that said label is oxidized and luminesces; and

35 (e) measuring the emitted luminescence to measure the presence of the analyte of interest in sample.

29. A method for performing a binding assay for an analyte of interest present in a sample comprising the steps of:

- 5 (a) forming a composition containing
  - (i) said sample;
  - (ii) an assay-performance-substance which contains a component linked to a label compound capable of chemiluminescing when oxidized; and
  - 10 (iii) a plurality of coated magnetic particles capable of specifically binding with the analyte and/or said assay-performance-substance;
- (b) introducing to said composition a molecule capable  
15 of being electrochemically converted to an oxidant capable of oxidizing said label;
- (c) incubating said composition to form a complex which includes a particle and said label compound;
- (d) magnetically collecting said complex at the  
20 surface of an electrode;
- (e) converting said molecule to said oxidant by imposing a voltage on said electrode, such that said label is oxidized and luminesces; and
- (f) measuring the emitted luminescence to measure the  
25 presence of the analyte of interest in sample.

30. A method for performing a binding assay for an analyte of interest present in sample comprising the steps of:

- 30 (a) forming a composition containing
  - (i) said sample;
  - (ii) an assay-performance-substance which contains a component linked to a label compound capable of chemiluminescing when oxidized; and
  - 35 (iii) a plurality of coated magnetic particles capable of specifically binding with the analyte and/or said assay-performance-substance;

(b) incubating said composition to form a complex which includes a particle and said label compound;

(c) introducing to said composition a molecule capable of being electrochemically converted to an oxidant capable  
5 of oxidizing said label;

(d) magnetically collecting said complex at the surface of an electrode;

(e) converting said molecule to said oxidant by imposing a voltage on said electrode, such that said label  
10 is oxidized and luminesces; and

(f) measuring the emitted luminescence to measure the presence of the analyte of interest in the sample.

31. A method for performing a binding assay for an analyte of interest present in sample comprising the  
15 steps of:

(a) forming a composition containing

(i) said sample;

(ii) an assay-performance-substance which contains a component linked to a label compound capable of chemiluminescing when oxidized;  
20 and

(iii) a plurality of coated magnetic particles capable of specifically binding with the analyte and/or said assay-performance-substance;  
25

(b) incubating said composition to form a complex which includes a particle and said label compound;

(c) magnetically collecting said complex at the surface of an electrode;

(d) introducing to said composition a molecule capable of being electrochemically converted to an oxidant capable of oxidizing said label;  
30

(e) converting said molecule to said oxidant by imposing a voltage on said electrode, such that said label  
35 is oxidized and luminesces; and

(f) measuring the emitted luminescence to measure the presence of the analyte of interest in the sample.

32. A method as recited in claim 22 conducted a batch process, the composition being permitted to reside within said cell for a time sufficient to permit collection of said particles.

5 33. A method as recited in claim 22 conducted as a flow process wherein said composition is flowed through said cell at a sufficiently low rate to permit collection of at least a portion of said particles.

34. An assay method as recited in claim 22  
10 wherein said particles have a density of from 0.1 to 5g/mL.

35. An assay method as recited in claim 34 wherein said particles have a density of from 0.5 to 2g/mL.

36. An assay method as recited in claim 22 wherein the size of said particles, measured as the mean  
15 diameter, ranges from 0.001 to 100  $\mu\text{m}$ .

37. An assay method as recited in claim 36 wherein the size of said particles ranges from 0.01 to 10  $\mu\text{m}$ .

38. An assay method as recited in claim 22  
20 wherein the concentration of particles in said composition is from 1 to 10,000  $\mu\text{g/mL}$ .

39. An assay method as recited in claim 38 wherein said concentration of particles is in the range of from 5 to 1000  $\mu\text{g/mL}$ .

25 40. A method as recited in claim 22 wherein said particles have a magnetic susceptibility is at least 0.001 cgs units.

41. A method as recited in claim 40 wherein the magnetic susceptibility is at least 0.01 cg units.

30 42. An assay method as recited in claim 22 wherein the magnetic susceptibility, density, size and concentration of said particles in said composition is such that the settling rate of said particles is at least 0.5 mm/min.

35 43. An apparatus for performing a binding assay for an analyte of interest present in sample based upon measurement of chemiluminescence comprising:

(a) a cell defining a sample containing volume having a vertical columnar zone having inlet and outlet means, and further including means for generating a magnetic field, and an electrode positioned below a substantial volume of said cell and said columnar zone; and

(b) means to impress a voltage upon said electrode; and

(c) means to measure the chemiluminescence generated at a collection zone.

44. An apparatus as recited in claim 43 wherein said means for generating a magnetic field includes plurality of magnets in north-south orientation and separated by non-magnetic material.

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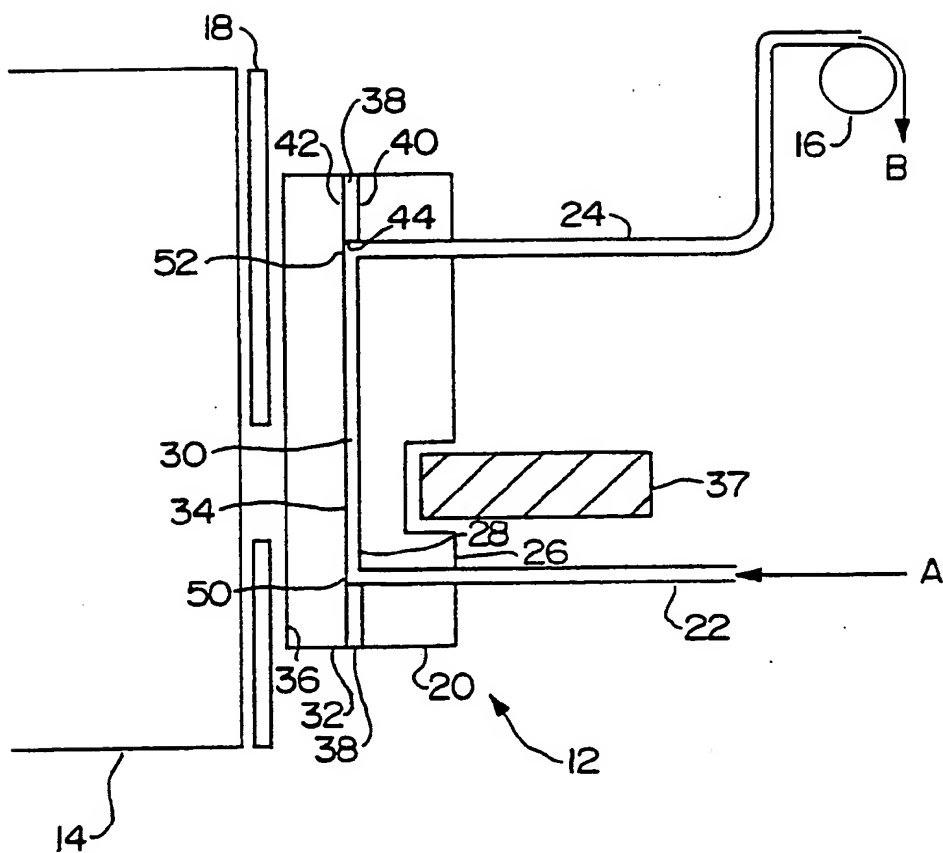
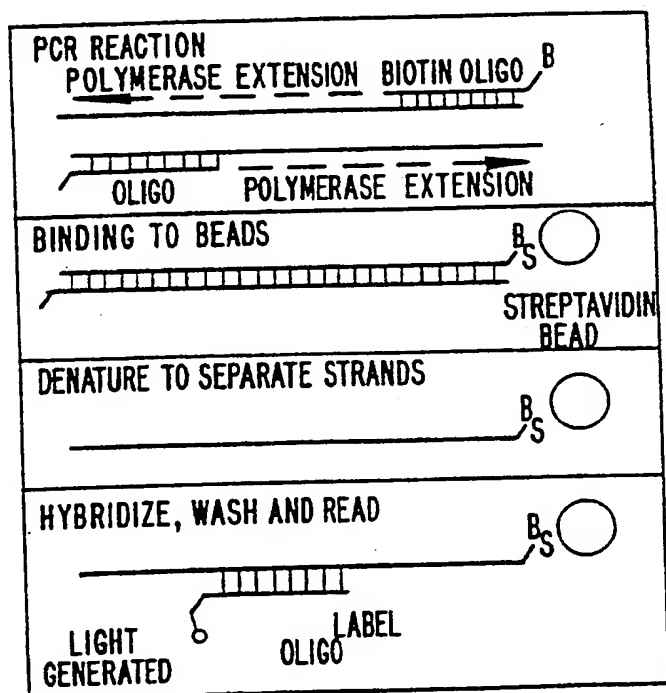
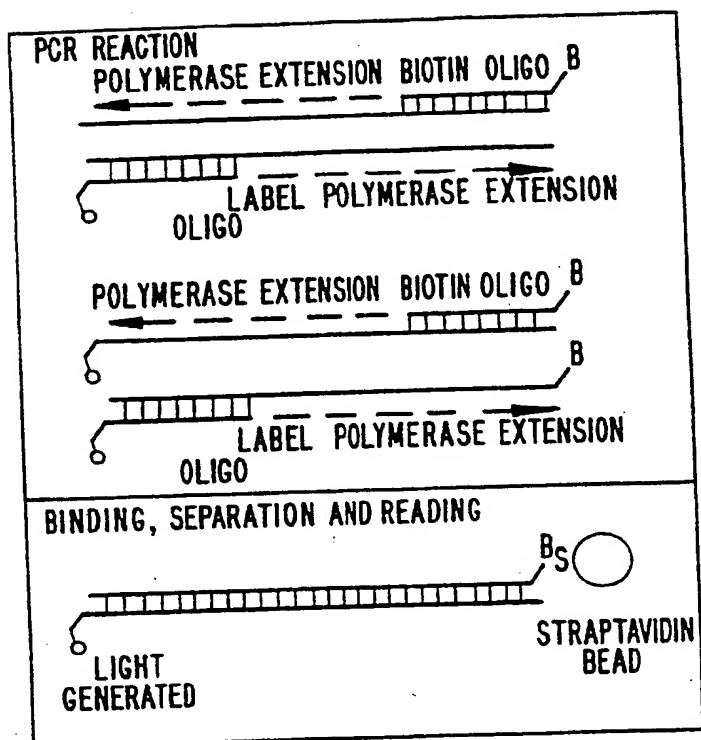
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FIG. 1

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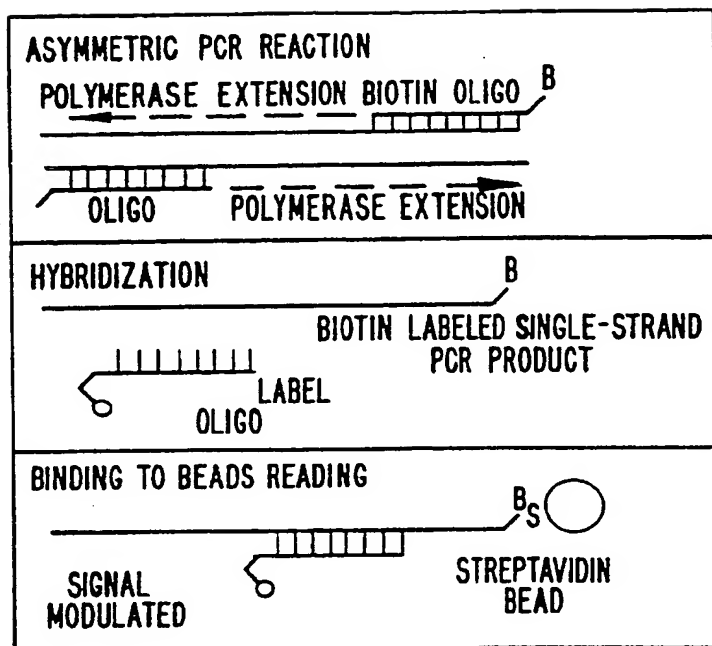


FIG. 4

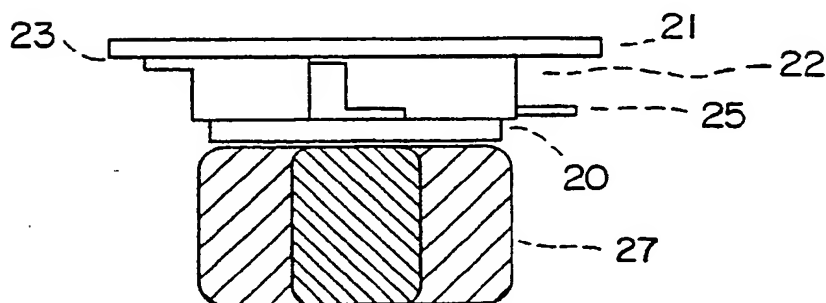


FIG. 5

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FIG. 6A

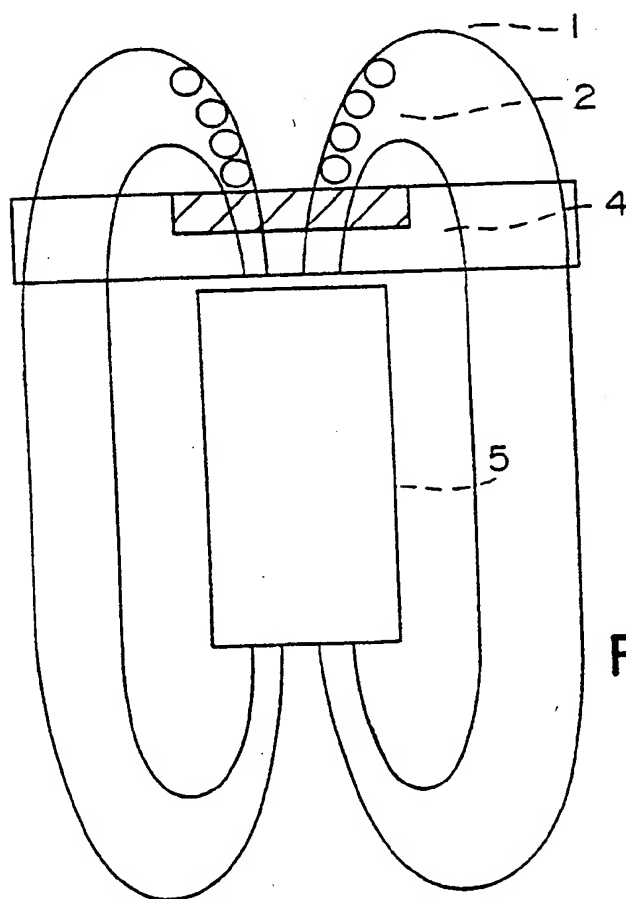
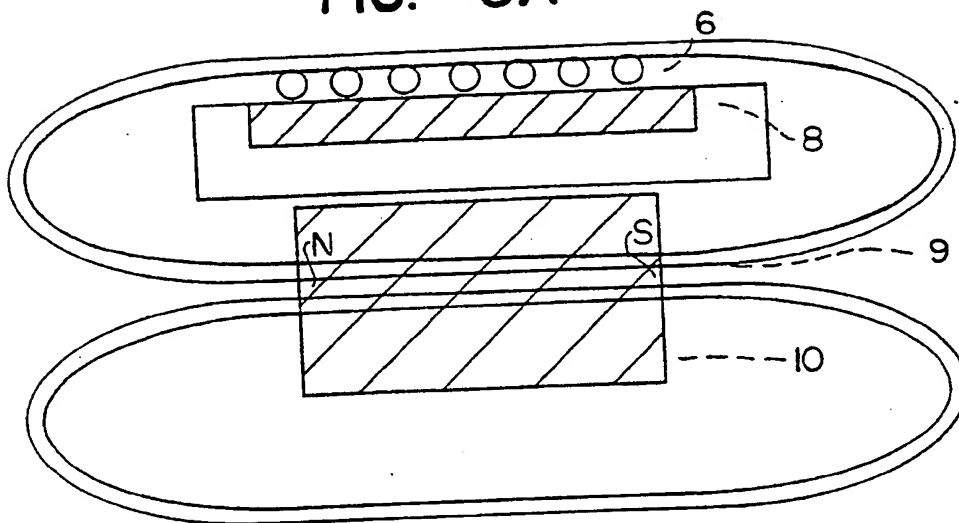


FIG. 6B

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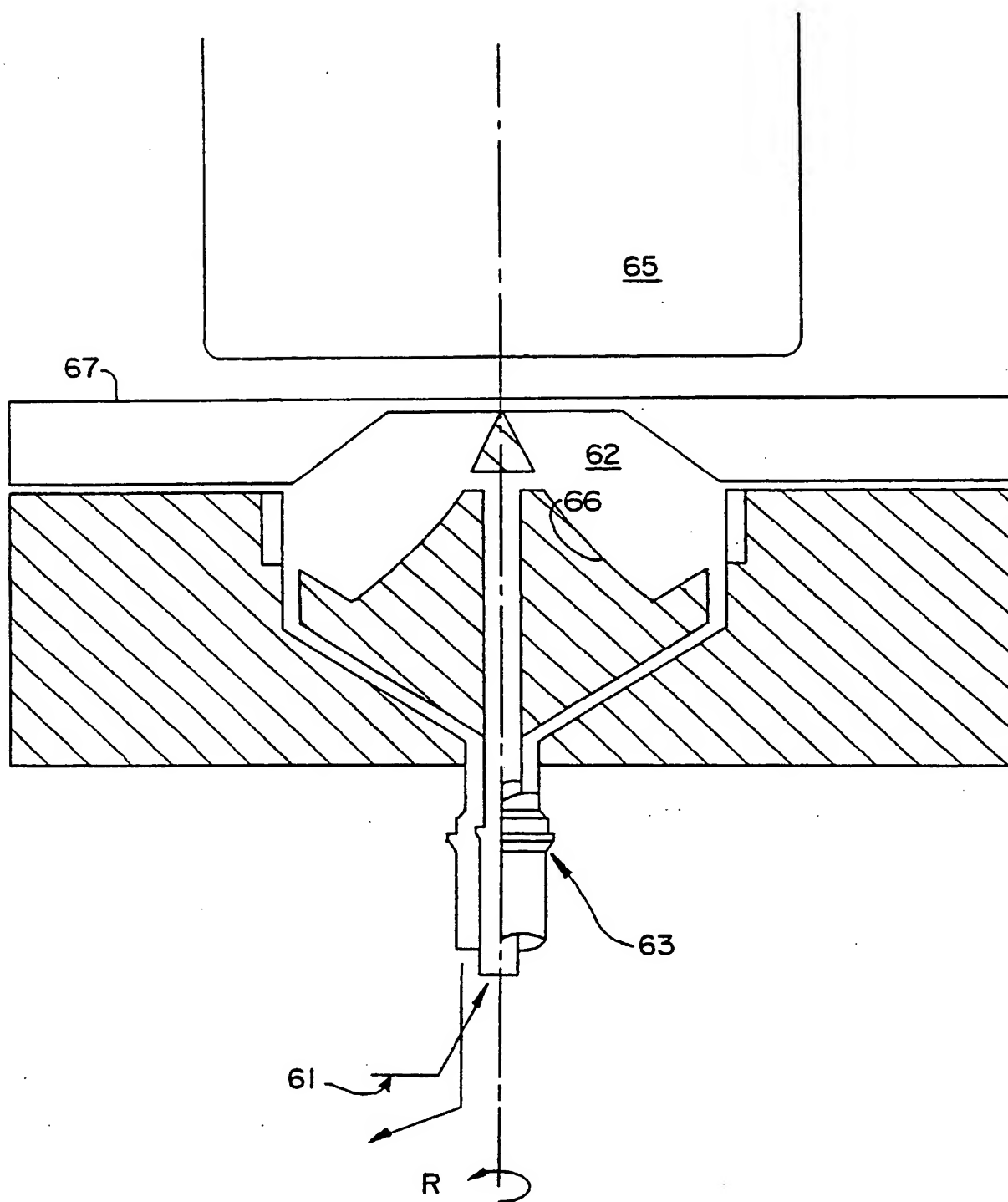
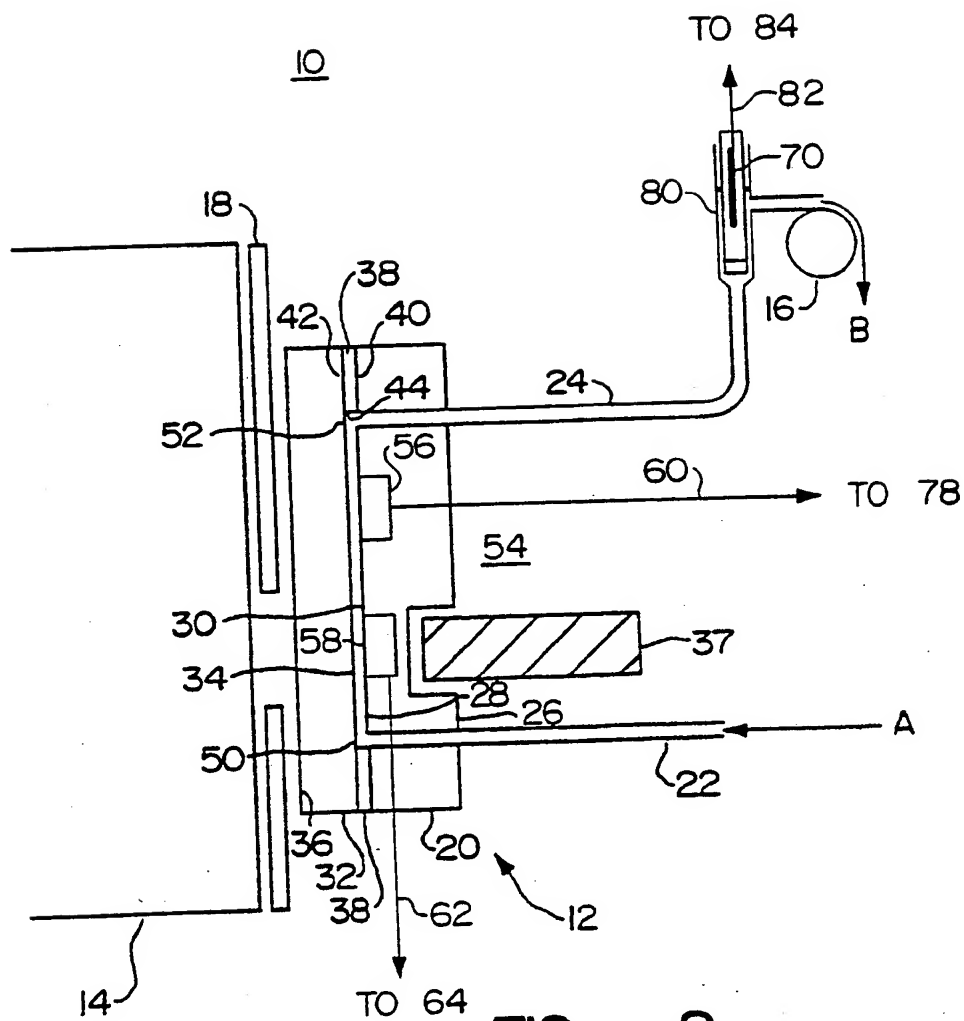
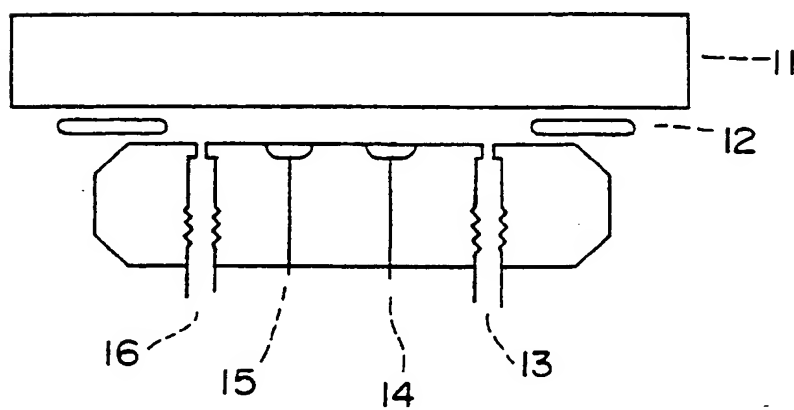
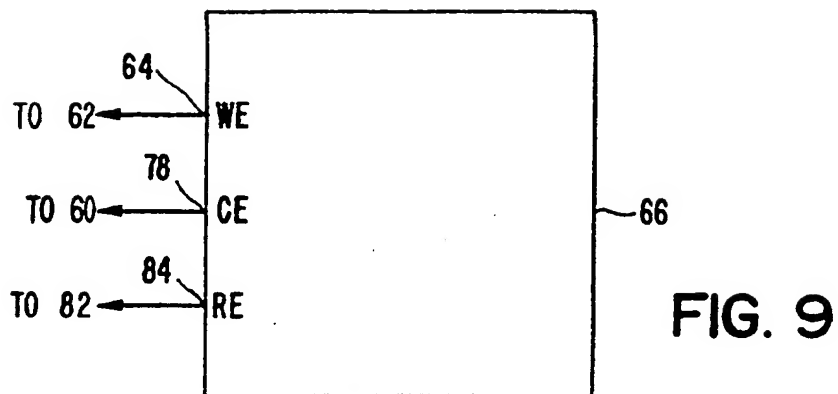


FIG. 7

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# INTERNATIONAL SEARCH REPORT

International application No.  
PCT/US92/05788

## A. CLASSIFICATION OF SUBJECT MATTER

IPC(5) :C12Q 1/28; G01N 33/533

US CL :435/7.1; 436/501

According to International Patent Classification (IPC) or to both national classification and IPC

## B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

U.S. : 435/7.1, 8; 436/501, 546

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practicable, search terms used)

APS, DIALOG

SEARCH TERMS ECL, ASSAY, MAGNETIC, HYBRIDIZATION, IMMUNOASSAY

## C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
Y	US, A, 4,628,037 (CHAGNON ET AL.) 09 DECEMBER 1986, SEE ENTIRE DOCUMENT.	1-44
Y,P	US, A, 5,068,088 (HALL ET AL.) 26 NOVEMBER 1991, SEE ENTIRE DOCUMENT.	20-21, 43-44
Y,P	US, A, 5,061,445 (ZOSKI ET AL.) 29 OCTOBER 1991, SEE ENTIRE DOCUMENT.	20-21, 43-44
Y	US, A, 4,280,815 (OBERHARDT ET AL.) 28 JULY 1981, SEE ENTIRE DOCUMENT.	1-19, 22-42
Y	US, A, 4,731,337 (LUOTOLA ET AL.) 15 MARCH 1988, SEE ENTIRE DOCUMENT.	1-19, 22-42
Y	US, A, 4,652,533 (JOLLEY ET AL.) 24 MARCH 1987, SEE ENTIRE DOCUMENT.	1-19, 22-42
Y	WO, A, 86/05815 (HILL ET AL.) 09 OCTOBER 1986, SEE ENTIRE DOCUMENT.	1-19, 22-42
Y	WO, A, 86/02734 (BARD ET AL.) 09 MAY 1986, SEE ENTIRE DOCUMENT.	1-19, 22-42
Y	WO, A, 87/06706 (MASSEY ET AL.) 05 NOVEMBER 1987, SEE ENTIRE DOCUMENT.	1-19,22-42

☒ Further documents are listed in the continuation of Box C. ☐ See patent family annex.

* Special categories of cited documents:	*T	later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention
*A* document defining the general state of the art which is not considered to be part of particular relevance	*X*	document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone
*E* earlier document published on or after the international filing date	*Y*	document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art
*L* document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)	*Z*	document member of the same patent family
*O* document referring to an oral disclosure, use, exhibition or other means		
*P* document published prior to the international filing date but later than the priority date claimed		

Date of the actual completion of the international search 16 October 1992	Date of mailing of the international search report OCT 30 1992
Name and mailing address of the ISA/ Commissioner of Patents and Trademarks Box PCT Washington, D.C. 20231 Facsimile No. NOT APPLICABLE	Authorized officer DAVID B. SCHMICKEL Telephone No. (703) 308-0196

Form PCT/ISA/210 (second sheet)(July 1992)\*

# INTERNATIONAL SEARCH REPORT

International application No.  
PCT/US92/05788

## C (Continuation). DOCUMENTS CONSIDERED TO BE RELEVANT

Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
A	BIOCHEM. BIOPHYS. RES. COMM. VOLUME 128, NO. 2, ISSUED 30 APRIL, 1985, Y. IKARIYAMA ET AL., "ELECTROCHEMICAL LUMINESCENCE-BASED HOMOGENEOUS IMMUNOASSAY", PAGES 987-992, SEE ENTIRE DOCUMENT.	1-44
Y,P	US, A, 5,093,268 (LEVENTIS ET AL) 03 MARCH 1992, SEE ENTIRE DOCUMENT.	20-21, 43-44

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